

# Mitochondrial Bioenergetics During Diapause in Embryos of *Artemia franciscana*

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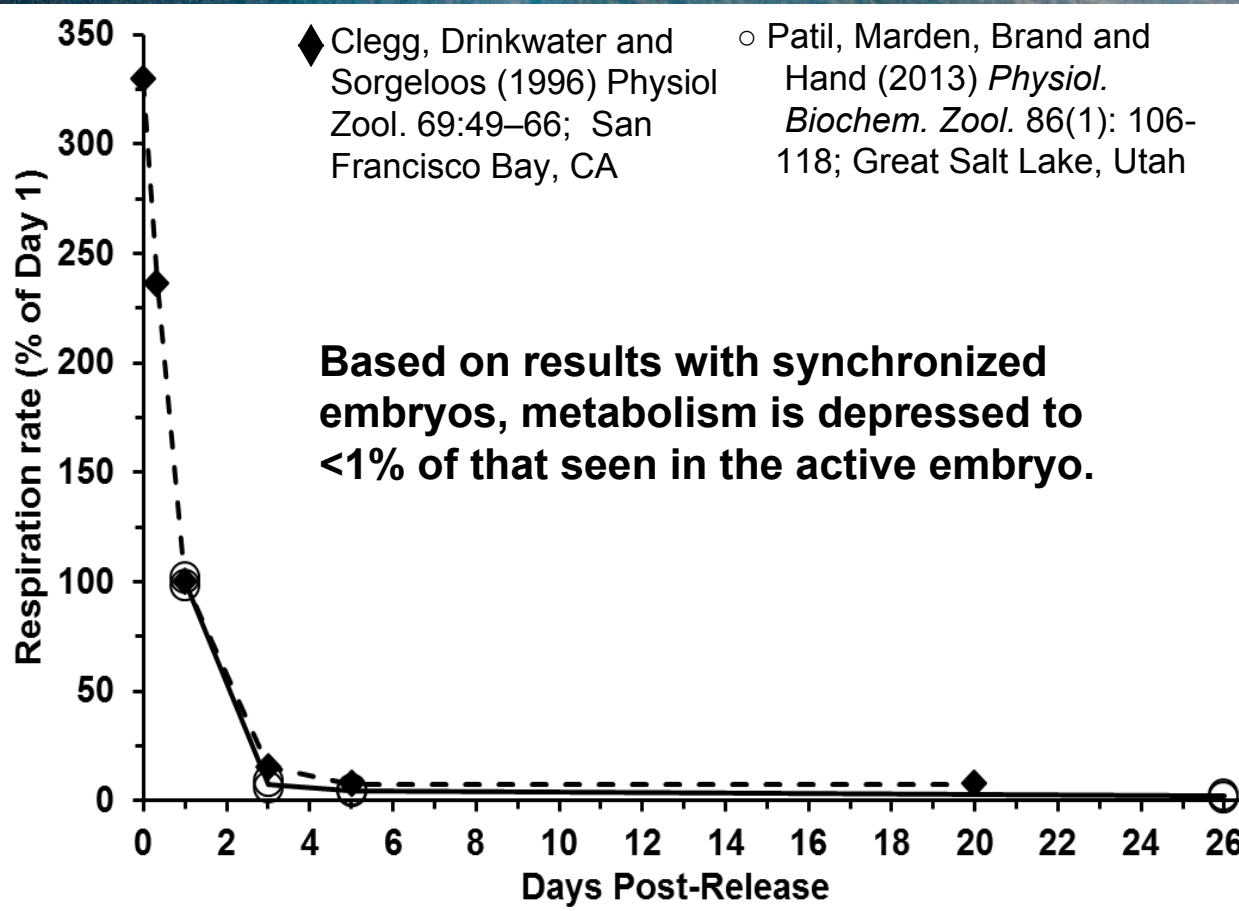
# Background and Aims of the Presentation

Diapause seen many invertebrates is a developmentally-programmed reduction of development and often metabolism, the depth of which can be profound – particularly in *Artemia franciscana*. Metabolic depression is positively correlated with extended survival during environmental stress.

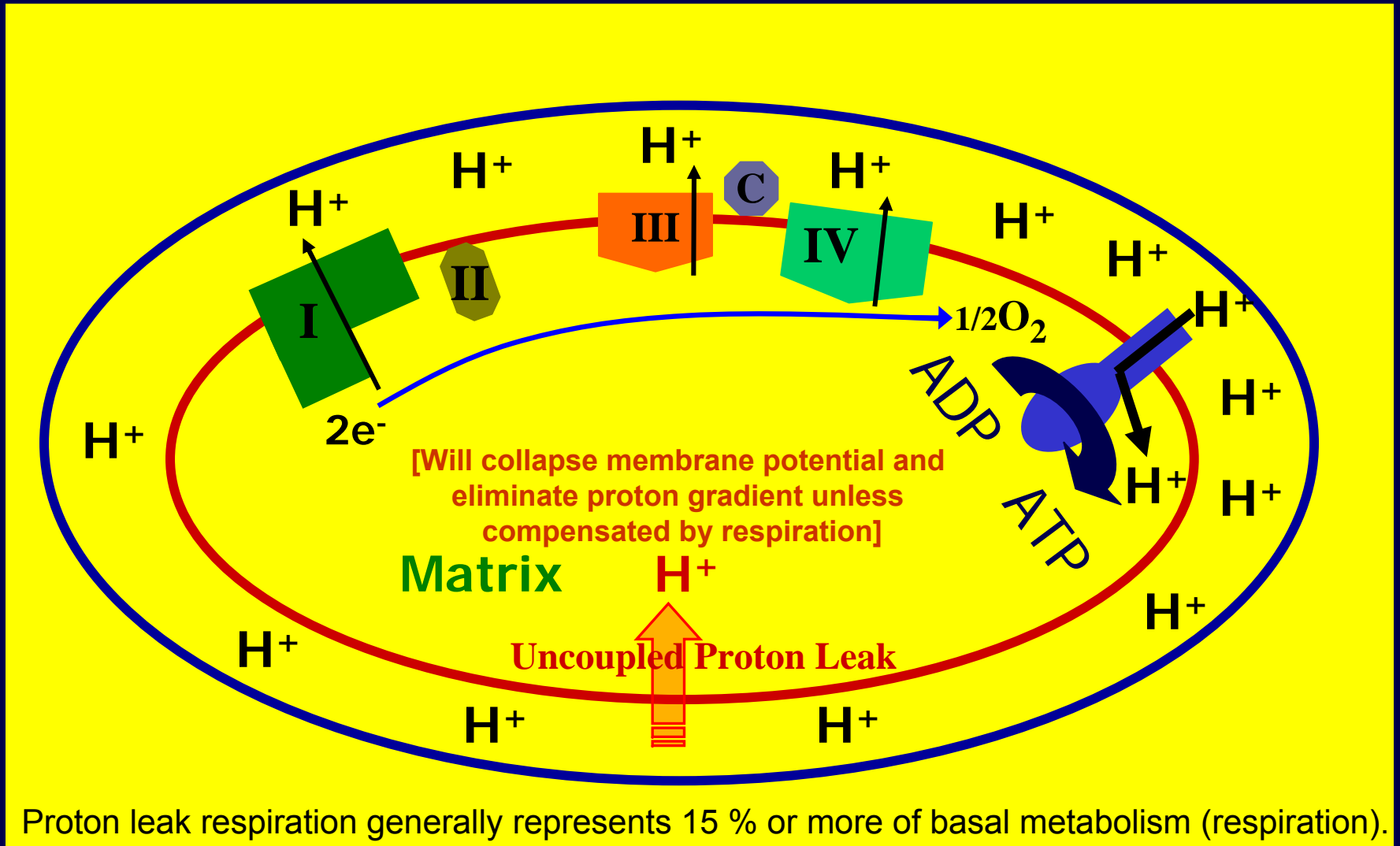
We have recently provided data suggesting the membrane potential ( $\Delta\Psi_m$ ) across the inner mitochondrial membrane is compromised during diapause, which has major implications for [1] cellular ATP status and [2] cell death pathways.

Knowledge of the complete sequence identities for the 22 or so subunits of the *Artemia* ATP synthase and the Inhibitor of F<sub>1</sub> Protein (IF<sub>1</sub>) would advance our understanding of the above physiological aspects of diapause. Genomic data are essential.

# Time Course for Respiratory Depression During Entry into Diapause



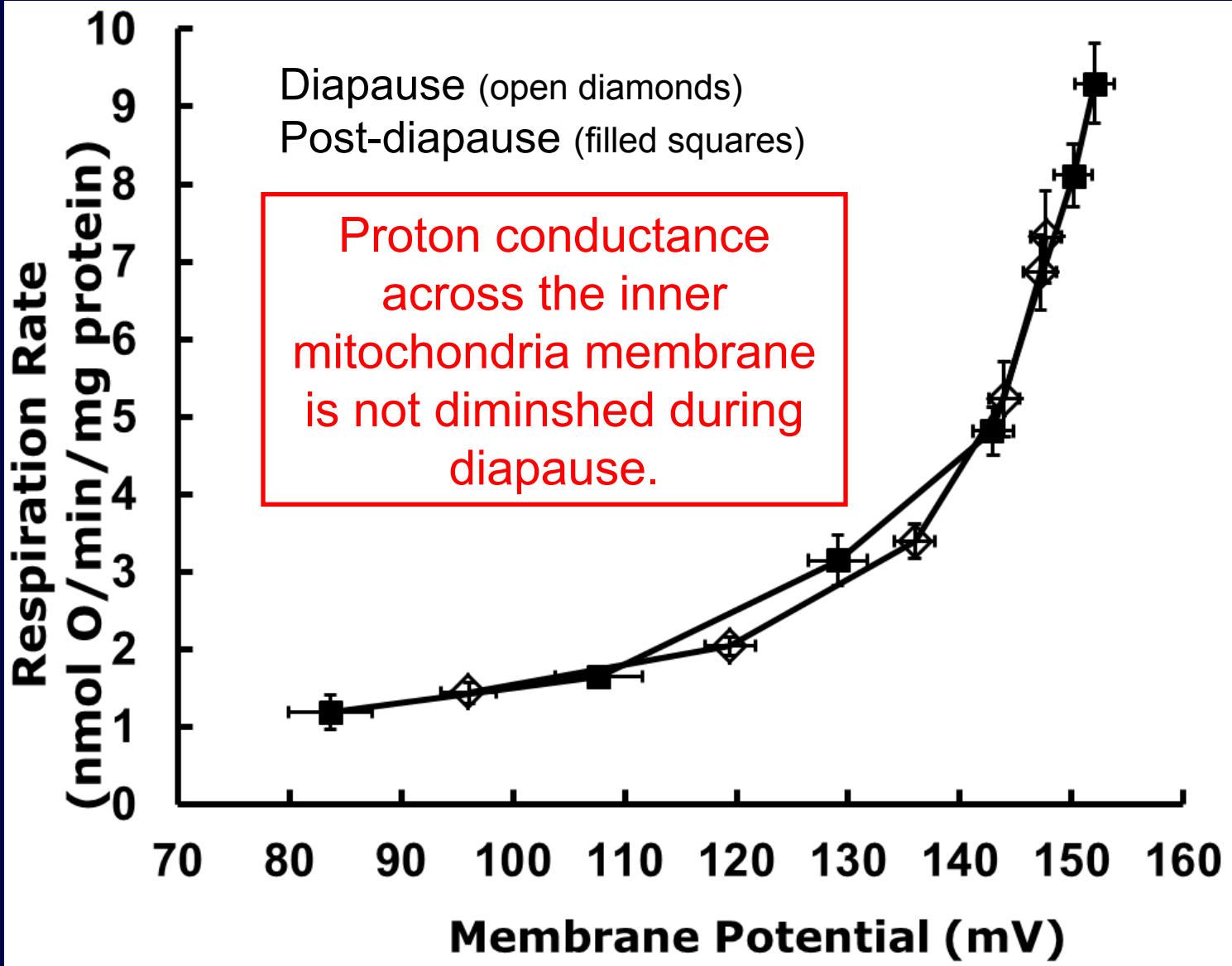
# Proton re-entry (leak) into the matrix and compensatory 'leak respiration'



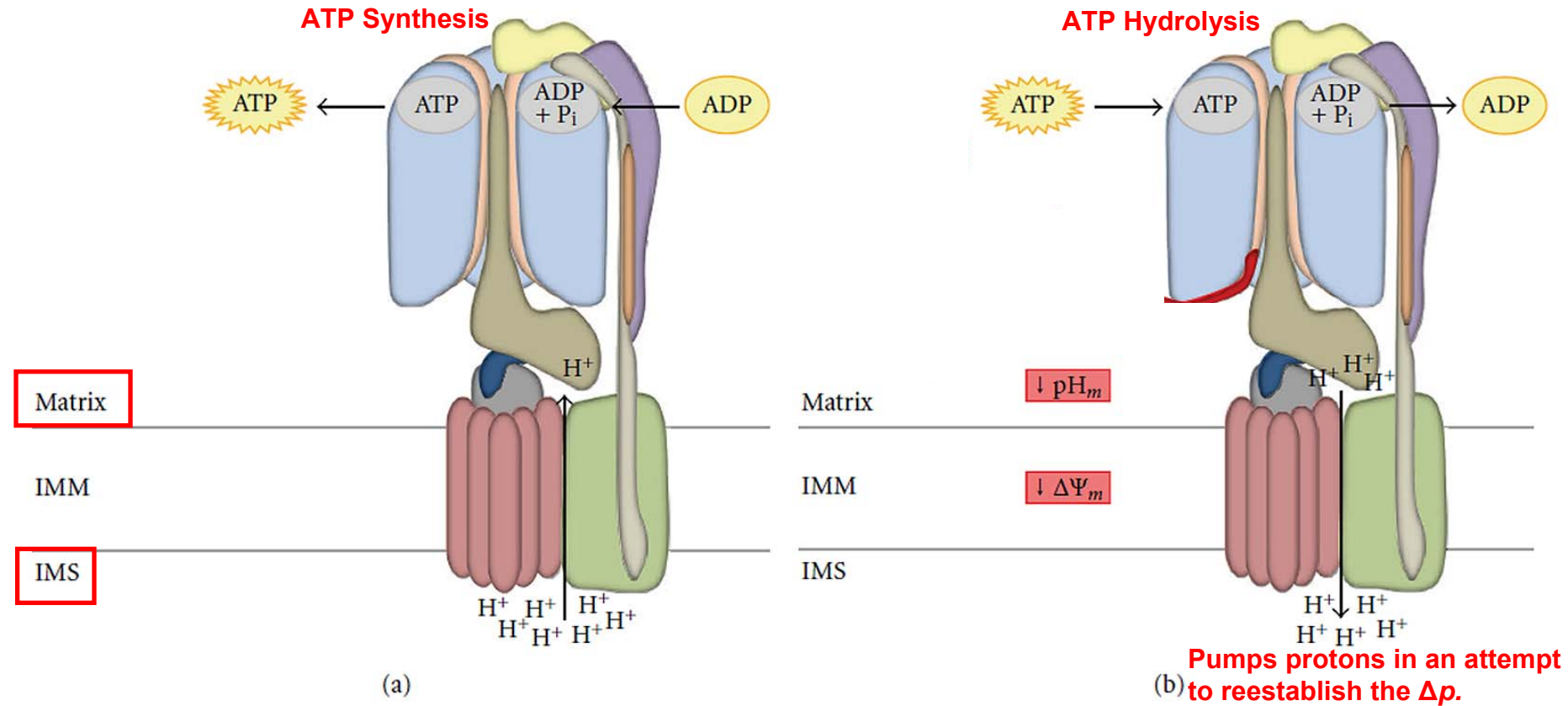
Proton leak respiration generally represents 15 % or more of basal metabolism (respiration).

# Proton Leak Respiration Versus Membrane Potential

Isolated Mitochondria from *Artemia franciscana* embryos



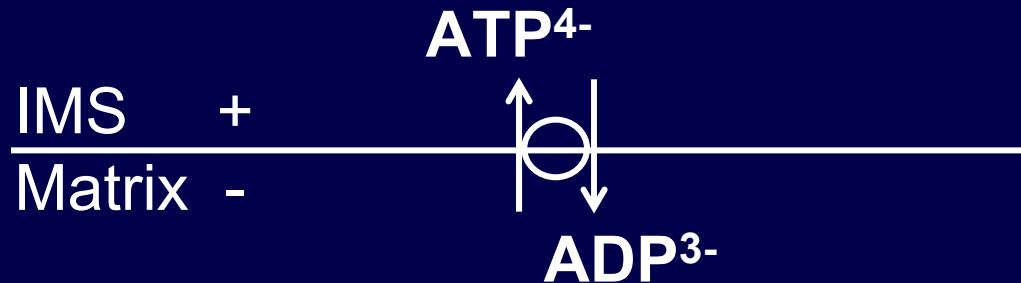
**Consequences?** Under such conditions of declining ATP, compromised  $\Delta\Psi$  and acidic matrix pH, the ATP synthase and the adenine nucleotide translocator (ANT) can reverse causing hydrolysis of all cellular ATP



[modified after: Faccenda D, Campanella M (2012) *Internat. J. Cell Biol.*, Article ID 367934, doi:10.1155/2012/367934]

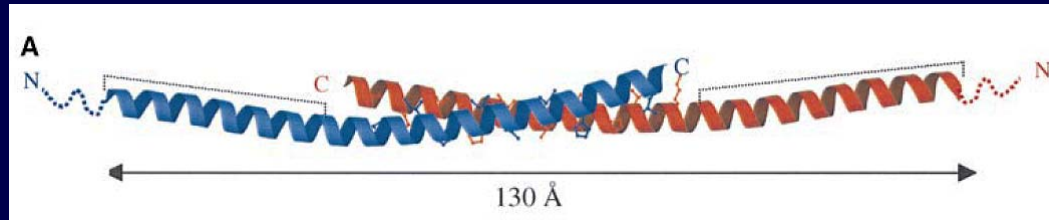
# The Adenine Nucleotide Translocator also Reverses

When there is a normal  $\Delta\Psi_m$ ,  $\text{ATP}_{\text{out}}/\text{ADP}_{\text{in}}$  is the favored direction for the ANT, because:



In the  $\Delta\Psi_m$  is compromised, there is no favoritism for the ANT – direction just depends on concentration gradient for ATP. **So ATP enters from the cytoplasm and can be readily cleaved by the ATP synthase, now functioning as a  $F_1F_0$  ATPase.**

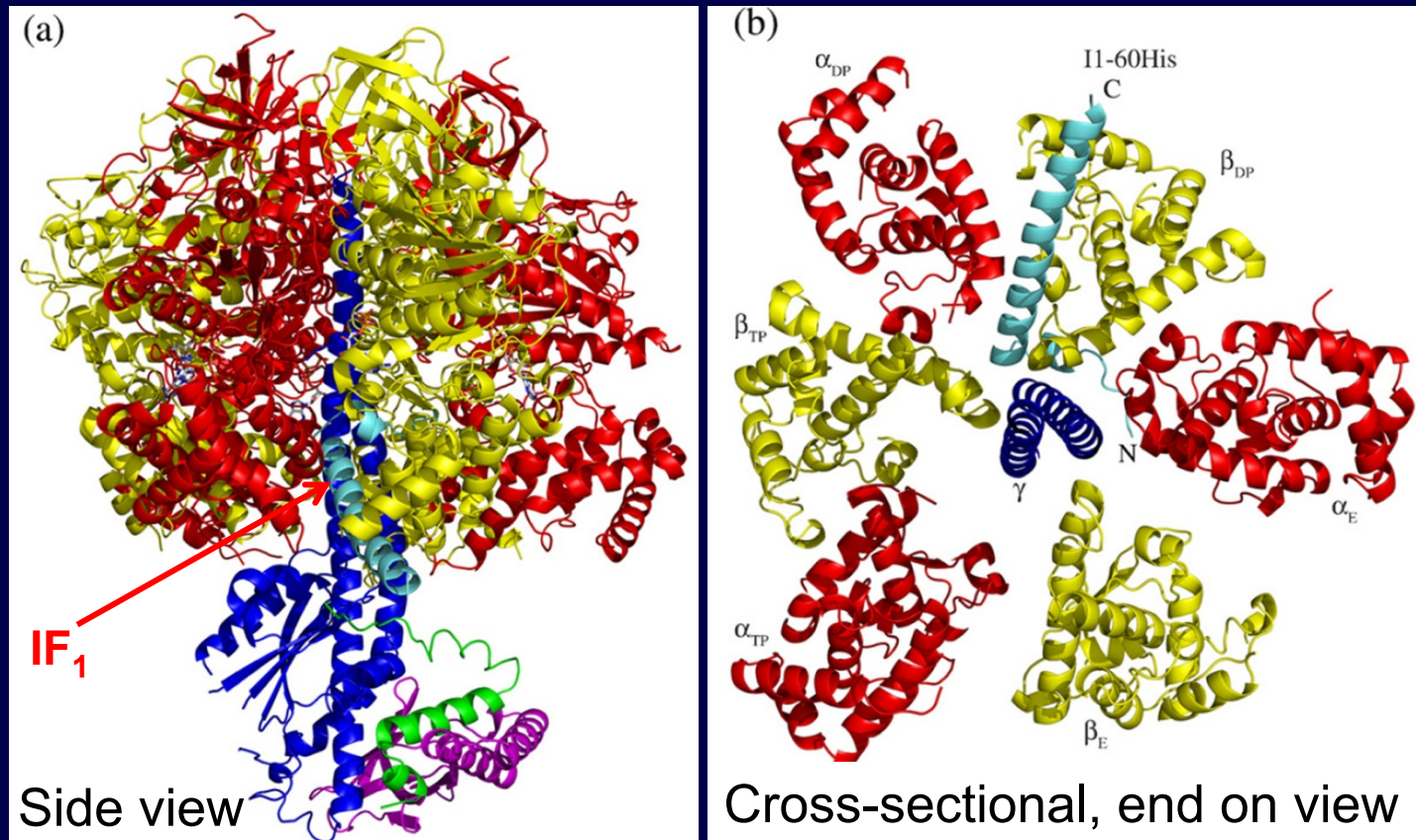
# Binding of IF<sub>1</sub> to Bovine ATP Synthase



Ribbon diagram of the *active dimer* of bovine IF<sub>1</sub> protein. Dashed lines represent the minimal inhibitory sequence. [from: Cabezon, Runswick, Leslie, Walker (2001) *EMBO J.* 20(24): 6990-6996]

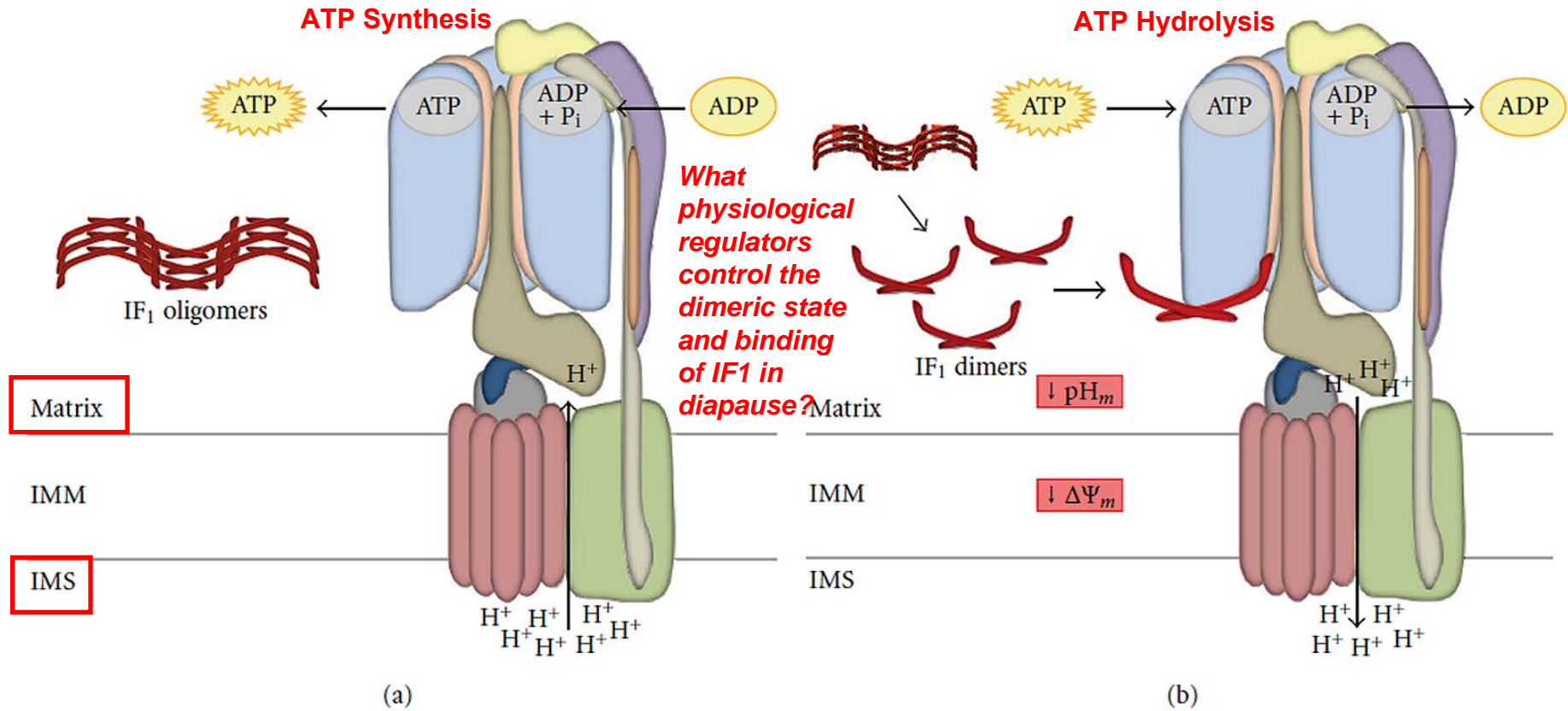
The structure of bovine F<sub>1</sub>-ATPase inhibited with residues 1–60 of the bovine inhibitor protein IF<sub>1</sub> (light blue). The inhibitor is bound at a catalytic interface between the  $\beta_{DP}$ - and  $\alpha_{DP}$ -subunits.

[modified from: Gledhill, Montgomery, Leslie, Walker (2007) *Proc. Natl. Acad. Sci. USA* 104 (40): 15671–15676]



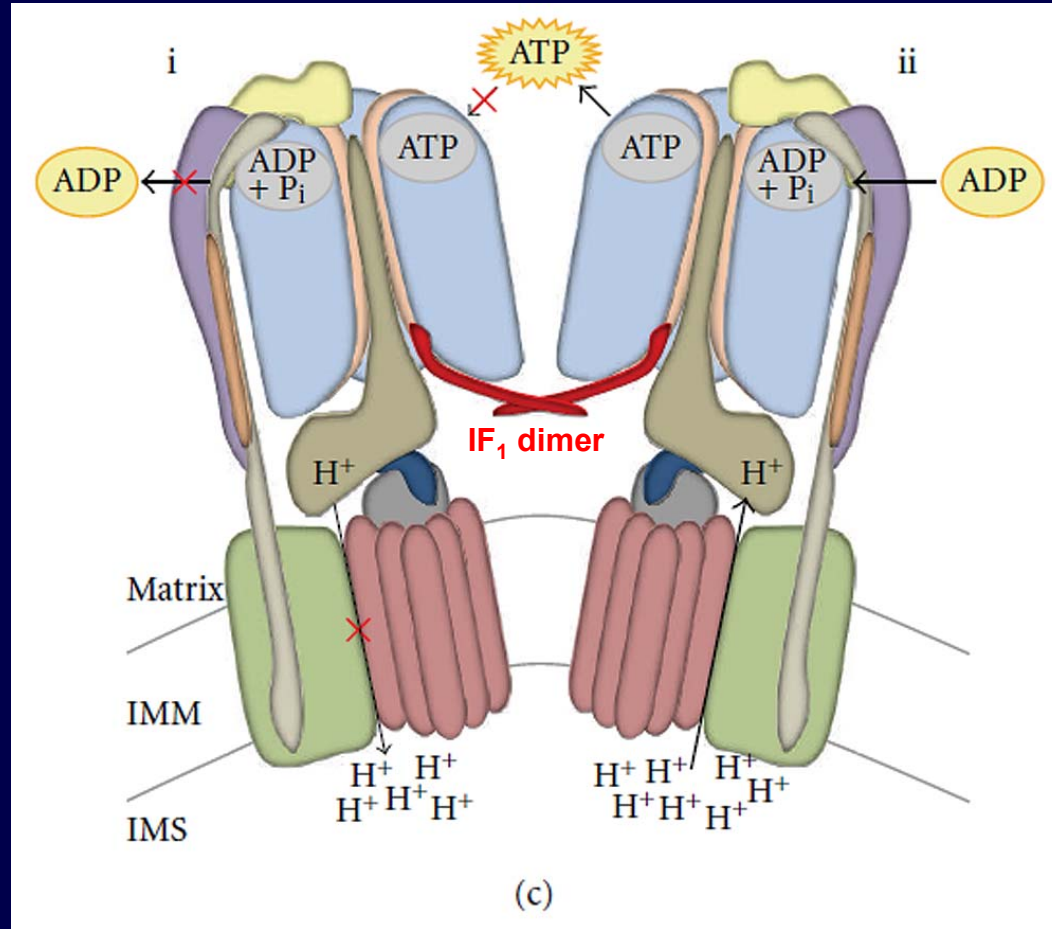


Normally  $IF_1$  exists as large oligomers, but these can depolymerize to free dimers that bind to the ATP synthase and inhibit hydrolysis



[modified after: Faccenda D, Campanella M (2012) *Internat. J. Cell Biol.*, Article ID 367934, doi:10.1155/2012/367934]

# ATP Synthase Inhibited by IF1



[Faccenda D, Campanella M (2012) *Internat. J. Cell Biol.* Article ID 367934, doi:10.1155/2012/367934]

Kinetically characterization of the interaction of IF<sub>1</sub> and the synthase in *Artemia* embryos requires the sequence of *Artemia* IF<sub>1</sub> protein so that the recombinant protein can be prepared.

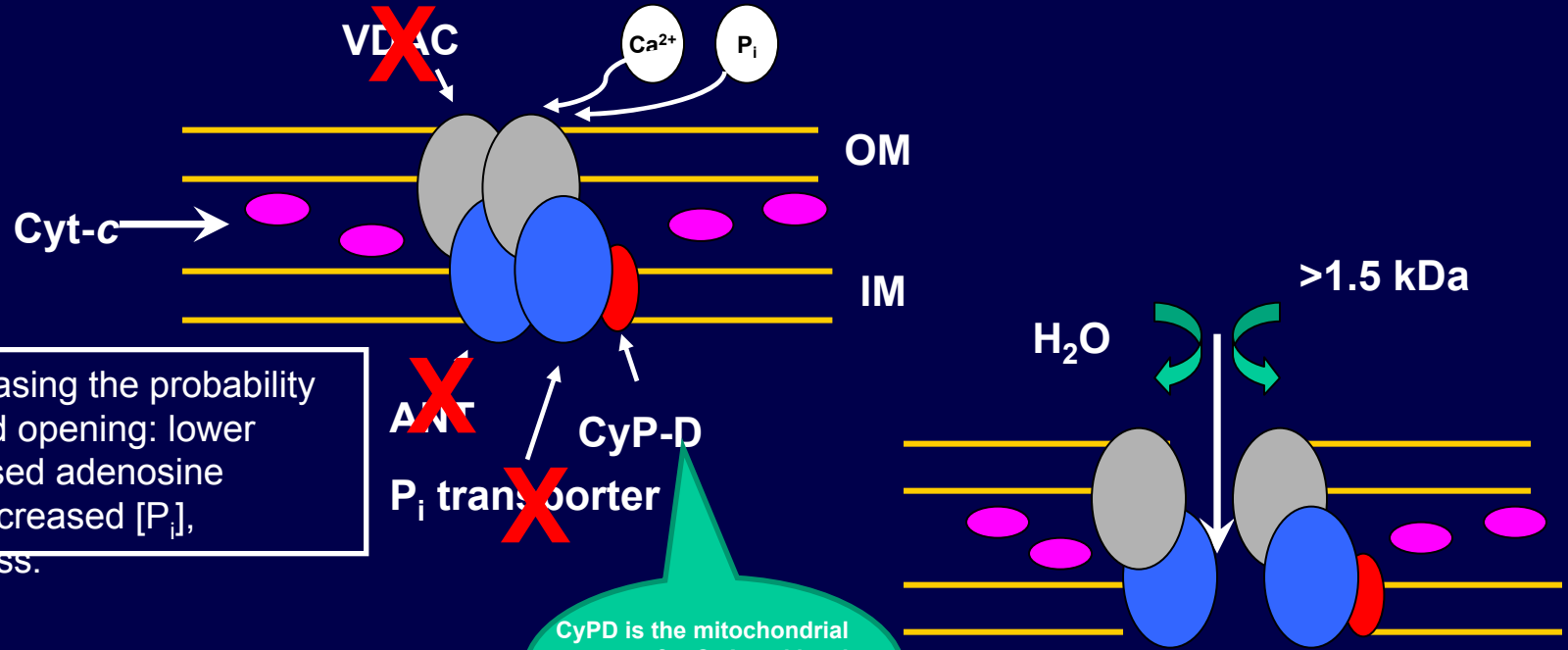
A second consequence of a disrupted mitochondrial  $\Delta\Psi_m$ ?

**Life is pleasant. Death is peaceful. It's the transition that is troublesome.**

**Isaac Asimov (January 2, 1920 – April 6, 1992)**



**MPTP:** A voltage dependent, cyclosporin A sensitive, and calcium-induced inner membrane channel with a ~1500 Da molecular weight cut off (Bernardi, 1992)

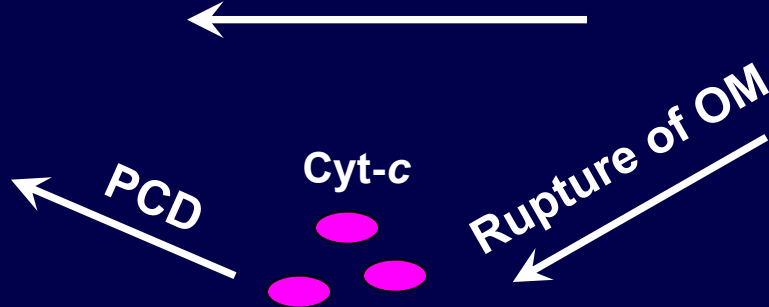


Factors increasing the probability of Ca-induced opening: lower  $\Delta\Psi_m$ , decreased adenosine nucleotide, increased  $[P_i]$ , oxidative stress.

CyPD is the mitochondrial receptor for CsA and is a key modulator of the MPTP, but not a structural component.



**Necrosis**

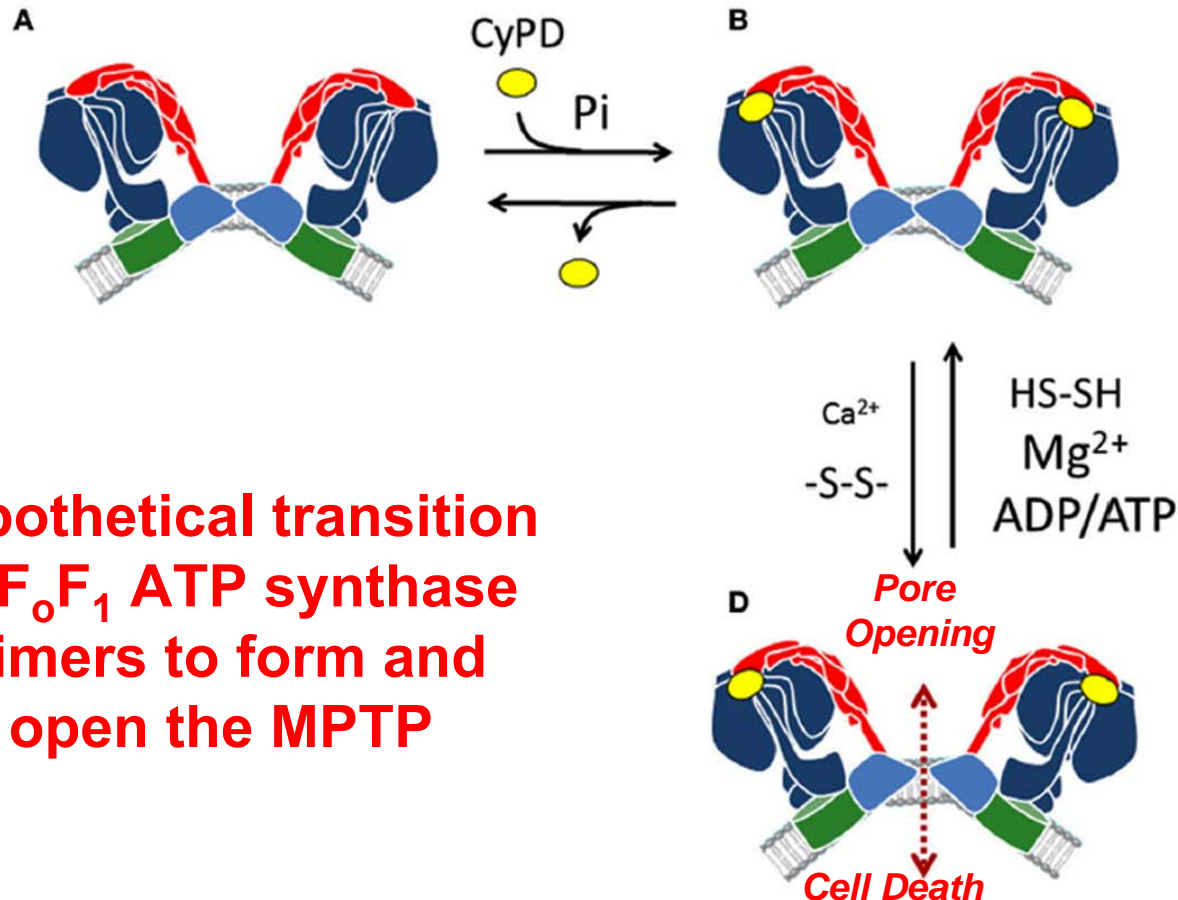


We have previously documented that the MPTP does not open in *A. franciscana* embryos in response to known physiological inducers or strong artificial inducers of pore opening.

[Menze, Hutchinson, Laborde, and Hand (2005) *Amer. J. Physiol.* 289: R68-R76]

*What is the basis for the absence of this phenomenon in A. franciscana?*

# Recent Evidence Suggests the MPTP is Formed by an Altered Conformation of the Dimeric ATP Synthase



**Hypothetical transition  
of F<sub>0</sub>F<sub>1</sub> ATP synthase  
dimers to form and  
open the MPTP**

Modified after: Giorgio et al (2013) *Proc. Natl. Acad. Sci. USA* 110: 5887–5892;  
Bernardi (2013) *Frontiers in Physiology* 4 doi: 10.3389/fphysiol.2013.00095

What structural/functional differences exist in the ATP Synthase from *A. franciscana* embryos that preclude opening of the MPTP?

Identification of subunits with genomic sequence data and comparison to the mammalian enzymes are important for resolving the MPTP issue in *A. franciscana* and would improve our understanding cell death and its prevention in general.

Additionally, the mechanisms for mitochondrial outer membrane permeabilization (involving Bcl-2 family proteins, Bax, Bak, etc.) need resolution in *Artemia* embryos.

*Genomic data for Artemia are key for advancing this work.*

# Lab Group, Collaborators, Support

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Mehmet Toner and colleagues (Harvard Med. School/MIT)

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